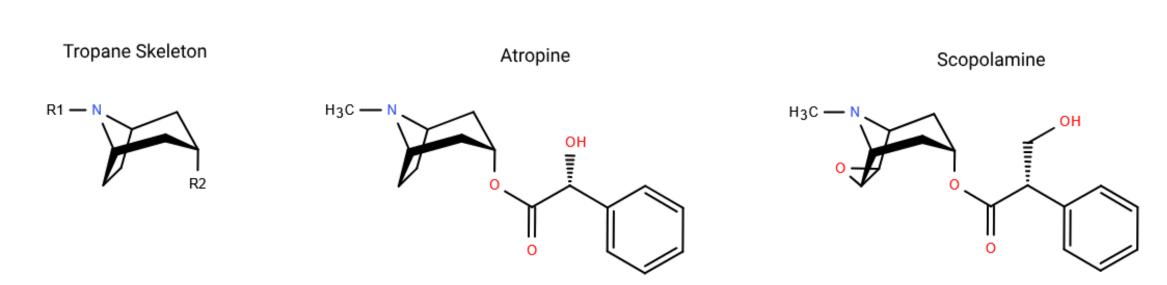
# Tropane Alkaloid Analysis Using SPE Combined With LC-MS/MS



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### Introduction

Tropane alkaloids are toxic compounds and are most abundant in the Solanaceae family of plants (deadly nightshade, henbane, mandrake and Jimson weed). Atropine and scopolamine are the most common tropane alkaloids found in food samples.<sup>2</sup> These alkaloids can contaminate herbal teas through plant debris<sup>3</sup> or soil migration.<sup>4</sup> They cause symptoms of reduced salivation, skin dryness, pupil dilation and with higher doses drowsiness, visual disturbances, palpitations, disorientation and hallucinations. The European Union (EU) has set a regulatory limit of 0.2 ng/mL<sup>6</sup> for the sum of atropine and scopolamine in herbal infusions.



 $\cdot$  Chemical structures of tropane alkaloids, including the tropane bicyclic ring structure, atropine and s

Current methods for analysis of tropane alkaloids in food use large bed weight solid phase extraction (SPE) products (>150 mg),<sup>1</sup> which are time consuming and less sustainable due to large solvent usage. This study validates the extraction of atropine and scopolamine with a 10 mg polymeric SCX 96 well SPE plate followed by HPLC-MS/MS analysis.

Following validation of the method, a range of herbal teas (chamomile, peppermint and green tea) sold on the UK market were analysed to assess the presence of atropine and scopolamine within the final herbal tea infusions.

# **Materials and Methods**

#### Tea infusion, according to ISO 3103:1980<sup>7</sup>:

2.00 g of tea was added to a stainless-steel tea strainer and placed in a beaker. 150 mL of boiling ultrapure water was poured through the tea strainer into the beaker and steeped for five minutes. The final tea solution was allowed to cool to room temperature and then filtered through a  $0.22 \mu m$  PES filter.

#### **SPE Method:**

A 10 mg 96 well plate (Microlute CP 10 mg SCX) with a positive pressure manifold (UltraPPM Lite) was used for processing samples.

- Conditioning: 1 mL of methanol
- Equilibration: 1 mL of 0.1% formic acid in ultrapure water
- Loading: 1 mL of acidified tea infusion sample (0.1% formic acid)
- Wash 1: 1 mL of 0.1% formic acid in ultrapure water
- Wash 2: 1 mL of 0.1% formic acid in methanol
- Drying step: Dried at 20 PSI for 2 minutes with the positive pressure manifold
- Elution: 2 x 0.5 mL of 0.5% ammonia in methanol
- Reconstitution: Eluate evaporated using a nitrogen blowdown evaporator (Ultravap Mistral) at 30°C and reconstituted with 0.2 mL of 0.1% formic acid in ultrapure water.

#### HPLC-MS/MS Method:

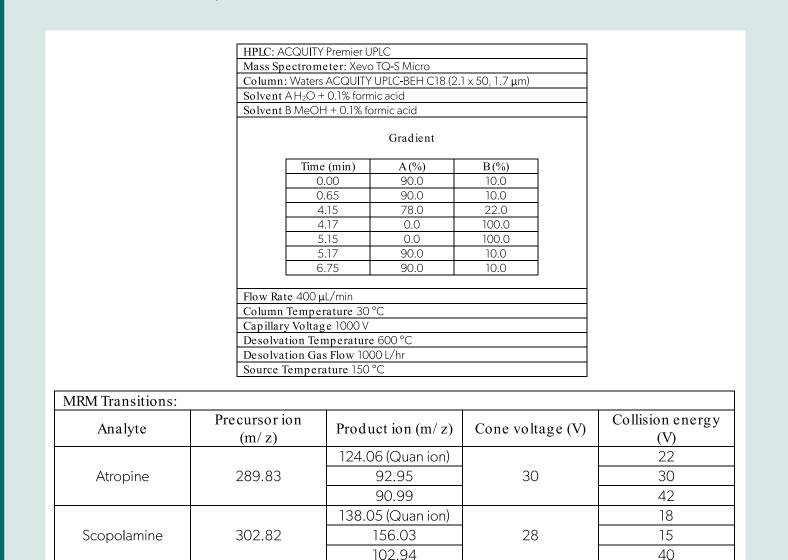


Table 1 - The HPLC and Mass Spectrometer conditions used for analysis and the atropine and scopolamine MRM transitions.

#### Validation guidelines:

Testing followed pesticide validation guidelines for food - SANTE/11312/20218 and Limit of Quantification was set by Regulation 2023/27839:

- **Recovery:** 70-120% at three concentrations (0.2, 1.0 and 5.0 ng/mL)
- **Repeatability:** ≤20% at each level
- Linearity:  $R^2 \ge 0.99$  with residuals  $\le \pm 20\%$
- Limit of Quantification (LOQ): <0.05 ng/mL</li>

### **Results and Discussion**

#### **Method validation – Linearity and Matrix Effects**

- Good linearity was achieved for all herbal infusions ( $R^2 = 0.991-0.996$ ) across 0.5-50 ng/mL range
- Low limits of quantification: 0.010 ng/mL for chamomile and peppermint, 0.025 ng/mL for green tea
- Negative matrix effects were observed for all samples (-12% to -38%)

Te a In fusion	Linearity (ng/ mL)	Atropine			Scopolamine		
		$\mathbb{R}^2$	LOQ (ng/mL)	ME (%)	$\mathbb{R}^2$	LOQ (ng/mL)	ME (%)
Chamomile	0.5 - 50	0.992	0.010	-17	0.995	0.010	-38
Peppermint	0.5 - 50	0.996	0.010	-14	0.992	0.010	-24
Green tea	0.5 - 50	0.991	0.025	-12	0.993	0.025	-22

Table 2 – Linearity, LOQ and matrix effects for chamomile, peppermint and green tea infusions, following SPE and HPLC-MS/MS analysis

#### Method validation – Recovery and Repeatability

- Matrix matched calibration used due to matrix effects > 20%
- High recovery rates achieved: 78-99% for both atropine and scopolamine across all matrices
- Excellent intra-day repeatability: 0.7-3.6%RSD
- Good inter-day repeatability: 2.1-15.8%RSD

	Atropine			Scopolamine			
Tea Infusion Sample	Recovery (%± SD)	Intra-day Repeatability (%RSD)	Inter-day Repeatability (%RSD)	Recovery (%± SD)	Intra-day Repeatability (%RSD)	Inter-day Repeatability (%RSD)	
Chamomile - 0.2 ng/mL	93 ± 1.7	1.8	6.9	84 ± 3.1	3.6	4.8	
Chamomile - 1 ng/mL	99 ± 1.6	1.6	9.8	94 ± 2.4	2.6	5.5	
Chamomile - 5 ng/mL	98 ± 2.0	2.1	2.4	99 ± 2.1	2.2	2.5	
Peppermint - 0.2 ng/mL	87 ± 0.88	1.0	2.4	85 ± 1.8	2.1	4.1	
Peppermint - 1 ng/mL	95 ± 0.78	0.8	4.5	96 ± 1.6	1.7	5.7	
Peppermint - 5 ng/mL	92 ± 1.6	1.7	2.2	95 ± 1.2	1.3	2.6	
Green Tea - 0.2 ng/mL	78 ± 2.7	3.4	15.8	78 ± 2.5	3.2	10.8	
Green Tea - 1 ng/mL	96 ± 1.2	1.3	5.1	93 ± 2.4	2.6	5.1	
Green Tea - 5 ng/mL	96 ± 0.66	0.7	3.9	96 ± 1.4	1.5	2.1	

Table 3 - Recovery and repeatability data for chamomile, peppermint and green tea infusions using the combined SPE and HPLC-MS/MS method for the analysis of atropine and scopolamine. Intra-day repeatability (n=8, in one day) and inter-day repeatability (n=16, on two days) SD = standard deviation

#### Application of the Method on Herbal Tea Infusions

- 12 commercial herbal tea samples were analysed (chamomile, peppermint, green tea)
- Most samples contained low or undetectable levels of atropine and scopolamine
- One chamomile sample (Chamomile-5) significantly exceeded EU regulatory limits:
  - Atropine: 0.59 ng/mL ( $\sim 3 \times EU$  limit of 0.2 ng/mL)
  - Scopolamine: 0.35 ng/mL (exceeds EU limit)
  - Sum of atropine and scopolamine: 0.94 ng/mL  $(\sim 5 \times EU limit)$

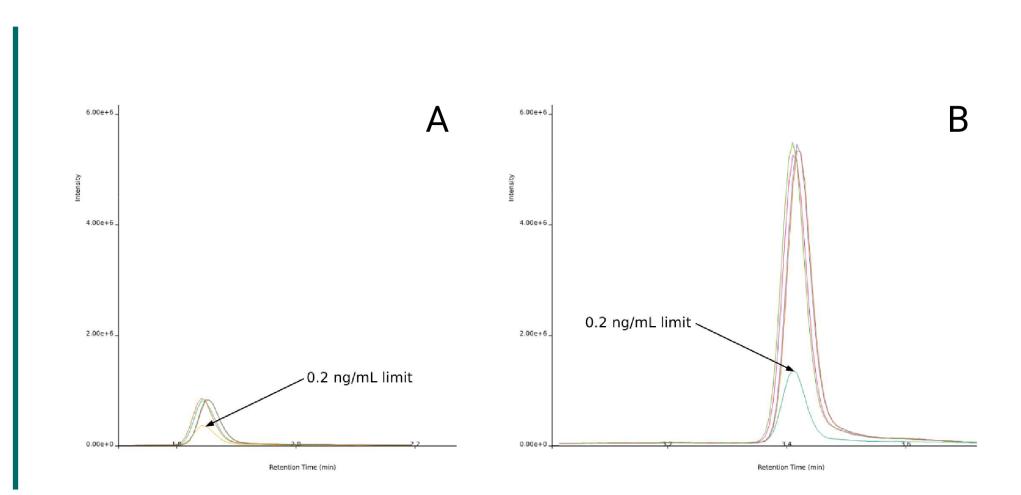


Figure 2 – Overlaid chromatograms showing the 0.2 ng/mL EU limit reference standard versus chamomile-5's chromatograms (n=4) at a mean concentration of 0.35 ng/mL for scopolamine (A) and 0.59 ng/mL for atropine (B).

Tea Infusion	Concentration (ng/mL)				
Sample	Atropine	Scopolamine	Sum of Atropine and Scopolamine		
Chamomile-1	<loq< td=""><td>ND</td><td><loq< td=""></loq<></td></loq<>	ND	<loq< td=""></loq<>		
Chamomile-2	<loq< td=""><td><loq< td=""><td><loq< td=""></loq<></td></loq<></td></loq<>	<loq< td=""><td><loq< td=""></loq<></td></loq<>	<loq< td=""></loq<>		
Chamomile-3	0.013 ± 0.0035	<loq< td=""><td>0.013 ± 0.0035</td></loq<>	0.013 ± 0.0035		
Chamomile-4	<loq< td=""><td>ND</td><td><loq< td=""></loq<></td></loq<>	ND	<loq< td=""></loq<>		
Chamomile-5	0.59 ± 0.0070	0.35 ± 0.012	0.94 ± 0.013		
Chamomile-6	<loq< td=""><td>ND</td><td><loq< td=""></loq<></td></loq<>	ND	<loq< td=""></loq<>		
Peppermint-1	<loq< td=""><td><loq< td=""><td><loq< td=""></loq<></td></loq<></td></loq<>	<loq< td=""><td><loq< td=""></loq<></td></loq<>	<loq< td=""></loq<>		
Peppermint-2	<loq< td=""><td><loq< td=""><td><loq< td=""></loq<></td></loq<></td></loq<>	<loq< td=""><td><loq< td=""></loq<></td></loq<>	<loq< td=""></loq<>		
Peppermint-3	<loq< td=""><td>0.011 ± 0.0014</td><td>0.011 ± 0.0014</td></loq<>	0.011 ± 0.0014	0.011 ± 0.0014		
Green tea-1	<loq< td=""><td>ND</td><td><loq< td=""></loq<></td></loq<>	ND	<loq< td=""></loq<>		
Green tea-2	ND	ND	ND		
Green tea-3	<loq< td=""><td>ND</td><td><loq< td=""></loq<></td></loq<>	ND	<loq< td=""></loq<>		

Table 4 - Concentration values (± standard deviation) for atropine, scopolamine and sum of both analytes in herbal infusions prepared using the SPE and HPLC-MS/MS method. ND = Not detected, due to signal to noise being less than 3:1

# Conclusions

- Validation was successful for the analysis of tropane alkaloids in herbal tea infusions using a 96-well SCX SPE method using a low bed weight (10 mg)
- The method meets SANTE/11312/2021 validation requirements with good linearity, low LOQs, and high recovery and good repeatability
- It provides a sustainable alternative to higher bed weight SPE methods
- Commercially available herbal teas generally contain low levels of tropane alkaloids
- However, one of the tested samples still contained a sample exceeding the EU limit showing there is still a risk to consumers

# References

1: L. González-Gómez, S. Morante-Zarcero, D. Pérez-Quintanilla, I. Sierra, Occurrence and Chemistry of Tropane Alkaloids in Foods, with a Focus on Sample Analysis Methods: A Review on Recent Trends and Technologica Advances, Foods, 2022, 11, 407, DOI: 10.3390/foods11030407.

2: P. P. J. Mulder, M. Nijs, M. Castellari, M. Hortos, S. MacDonald, C. Crews, J. Hajslova, M. Stranska, Occurrence of tropane alkaloids in food, EFSA Supporting Publications, 13, DOI: 10.2903/sp.efsa.2016.EN-1140.

3: M. Nijs, C. Crews, F. Dorgelo, S. MacDonald, P. P. J. Mulder, Emerging Issues on Tropane Alkaloid Contamination of Food in Europe, Toxins, 2023, 15, 98, DOI: 10.3390/toxins15020098 4: L. González-Gómez, S. Morante-Zarcero, J. A. M. Pereira, J. S. Câmara, I. Sierra, Improved Analytical Approach for Determination of Tropane Alkaloids in Leafy Vegetables Based on μ-QuEChERS Combined with HPLC-MS/MS,

Toxins (Basel), 2022, 14, 650, DOI: 10.3390/toxins14100650

5: Health for humans, animals & plants – Tropane Alkaloids, https://www.ages.at/en/human/nutrition-food/residues-contaminants-from-a-to-z/tropane-alkaloids, (accessed May 2025).

6: Commission Regulation (EU) 2023/915 of 25 April 2023 on maximum levels for certain contaminants in food and repealing Regulation (EC) No 1881/2006,

7: ISO 3103: Tea — Preparation of liquor for use in sensory tests, 1980 edition.

https://eur-lex.europa.eu/legal-content/EN/TXT/?uri=CELEX%3A02023R0915-20250101, (Accessed May 2025)

8: Analytical Quality Control And Method Validation Procedures For Pesticide Residues Analysis in Food and Feed - Sante 11312/2021 V2,

https://food.ec.europa.eu/system/files/2023-11/pesticides\_mrl\_guidelines\_wrkdoc\_2021-11312.pdf, (accessed April 2025).